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ACKNOWLEDGMENTS

Appreciation is expressed to Allen Ormiston of Amoco Production Company, Tulsa, Oklahoma, for information on the use of black opaque in fossil photography. J. G. Vedder kindly lent specimens of modern mollusks from the Gulf of California for use in illustrating this report.

The manuscript has been reviewed by William K. Emerson of the American Museum of Natural History, A. Myra Keen of Stanford University, David L. Jones and Warren O. Addicott of the U. S. Geological Survey, and their critical comments are deeply appreciated. Special thanks are due to Warren O. Addicott for his encouragement and advice on writing this report.

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Explanation of Figures 1 to 18

(all figures natural size)

Figures 1 to 4: Cassis sp., Baja California, Mexico. I untreated; 2 coated with dulling spray; 3 coated with ammonium chloride; 4 coated with black opaque and ammonium chloride.

Figures 5 to 8: Oliva sp., Baja California, Mexico. 5 untreated;

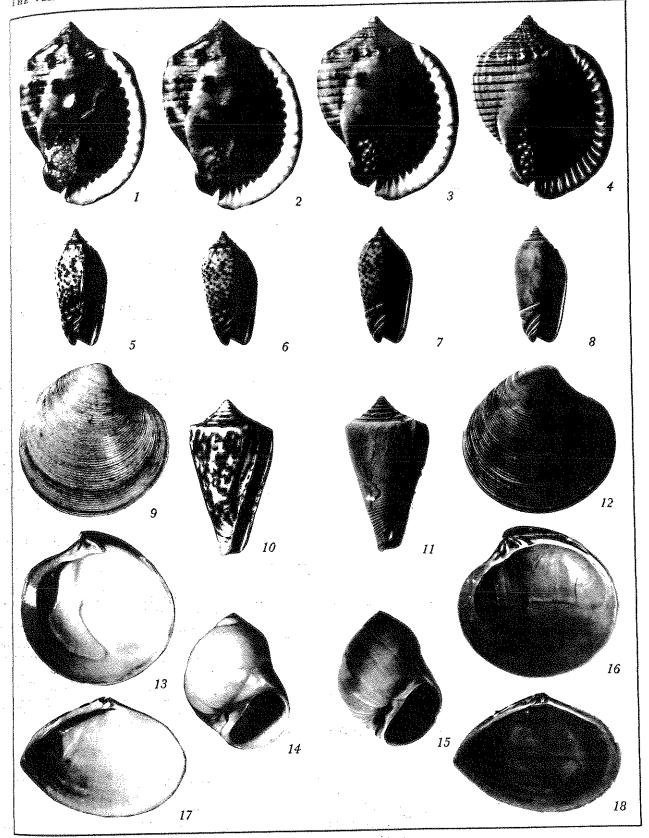
6 coated with dulling spray; 7 coated with ammonium chloride; 8 coated with black opaque and ammonium chloride.

Figures 9, 12, 13, and 16: Dosinia sp., Baja California, Mesco 9, 13, untreated; 12, 16, coated with black opaque and ammonium chloride.

Figures 10 and 11: Conus sp., Baja California, Mexico. 10 un treated; 11 coated with black opaque and ammonium chloride Figures 14 and 15: Polinices sp., Baja California, Mexico. 14

treated; 15 coated with black opaque and ammonium chloride. Figures 17 and 18: Macoma sp., Gulf of Alaska, Alaska. 17 un-

treated; 18 coated with black opaque and ammonium chloride.



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wigh searching from a permanently emplaced survey eases the difficulty in locating them. Other mollusks have been successfully tagged using method: The rock scallop, Hinnites multirugosus GALE, 1928); the red abalone, Haliotis rufescens; and * wasy top shell, Astraea undosa (Wood, 1828).

initial [] Kelletia kelletii were tagged on December

1967. Success in finding these motile benthonic ani-

is targely dependent upon the visibility underwater,

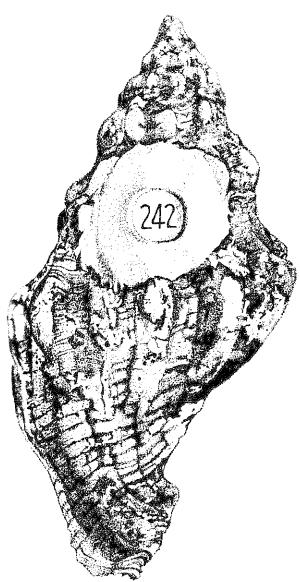


Figure 1

the drawing shows a tagged Kelletia kelletii with a numbered disc ssientided in epoxy and cemented to the dorsal region of the shell.

The size of the mollusk does not present a problem, since the epoxy can be shaped to conform to the characteristics of the outer shell and the disc tags sized accordingly. This reduces any hindrance to the movement of the animal and the possibility of the epoxy coming in contact with the animal's living tissue (Figure 1).

The advantages of this technique appear to be the permanence of marking, and the fact that tagging can be completely carried out underwater with a minimum amount of disturbance to the subjects and their environment.

ACKNOWLEDGMENTS

I am particularly grateful to Dr. W. D. Clarke for valuable advice and encouragement, and to Mr. R. E. Bower for technical assistance in the field.

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A Method of Color Preservation in Opisthobranch Mollusks

BY

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INTRODUCTION

Color and color pattern are useful taxonomic characters in the opisthobranch mollusks. However, I know of no published reports describing a method of preserving color in opisthobranchs. Instead, it is usually assumed that the colors will fade or be bleached out in formalin,

TOYAMA & MIYOSHI (1963) and Waller & Esch-MEYER (1965) successfully employed an antioxidant, butylated hydroxytoluene (hereafter referred to as Ionol C. P.-40), to preserve some colors in fish and a prawn. Waller & Eschmeyer (op. cit.) used 1, 10, and 20 cc of Ionol C. P.-40 per 4500 cc of 10% formalin to make the test solutions (0.02%, 0.22%, and 0.44% concentrations respectively). Eighteen months later, they found that the color of fish in the 0.44% solution was best preserved, but preservation of colors of fish in other test solutions was superior to that of fish in untreated formalin solution.

In this note, I report the results of testing this technique on opisthobranchs from 3 orders (Cephalaspidea, Sacoglossa, and Nudibranchia) with emphasis on the nudibranchs.

MATERIALS AND METHODS

A stable emulsion was formed by vigorously stirring Ionol C. P. -40 into hot sea water (55° to 65° C) (Shell Technical Bulletin, IC:67-16). Concentrations of Ionol C. P. -40 used in the emulsions gave final dilutions of 0.1% to 0.5% Ionol C. P. -40 by volume. Formalin was added to this emulsion to give a final concentration of 5% formalin by volume (hereafter referred to as Ionol C. P. -40 emulsion). Ionol C. P. -40 is also readily soluble in alcohol.

Most of the opisthobranchs were relaxed for 1 to 5 minutes, depending on size, with succinylcholine chloride (Beeman, 1968). A few, especially the Cephalaspidea, were relaxed for 2 to 8 hours in propylene phenoxetol using 1% by volume of propylene phenoxetol in sea water (Owen, 1955; Owen & Steedman, 1958).

Small animals (< 1.5 to 2.0 cm long) were then put directly into Ionol C. P. -40 emulsion. Formalin diffuses inward rapidly enough to preserve the internal tissues. Larger animals (> 2 cm long) were injected with a small amount of 5% formalin in sea water without Ionol C. P. -40, to preserve the internal tissues, then put into the Ionol C. P. -40 emulsion.

The Ionol C. P. -40 emulsion was not injected into the animal because it eventually forms an oily film in the body cavities and over the tissues. This is particularly undesirable if the animals are to be dissected. In any case, there are few internal organs in an opisthobranch in which color is distinctive or taxonomically important.

The method was tested on 1 sacoglossan, Elysia hedgpethi Marcus, 1961; 2 cephalaspideans, Haminoea sp., and Aglaja diomedea (Bergh, 1893); and 30 species of nudibranchs of which 12 were dorids, 12 were dendronotaceans, and 6 were acolids.

RESULTS

Of the concentrations tested, 0.3% Ionol C. P. -40 emulsion resulted in maximal color retention and was used for most of the specimens.

Stored in bottles in glass-door cabinets, the specimens in Ionol C. P. -40 emulsion have been exposed to normal artificial lighting, but protected from direct sunlight, for up to 2 years. In most cases, the original color has been retained albeit with varying degrees of fading in some species. The color of virtually all specimens stored under similar conditions, but in untreated 5% formalin in sea water, has faded markedly or disappeared.

Some colors are preserved better and longer than others. The yellow body color and black spots of Anusdoris nobilis (MACFARLAND, 1905) and Archidoris more terevensis (Cooper, 1862) are well preserved as are the yellow and orange pigments of Triopha carpenter (STEARNS, 1873). The orange on the cerata of Land cockerelli MacFarland, 1905 is completely preserved while the orange on Hermissenda crassicornis (Esch SCHOLTZ, 1831) faded slightly. Slight fading has occurred in the orange-red body color of Rostanga pulchra Moo FARLAND, 1905. The salmon-pink body color of Trionical festiva (Stearns, 1873) and T. gilberti (MACFARLAND 1966) and the orange body color of T. (Tochuina) terrs quetra (Pallas, 1788). Lade only slightly. The same > true of the brown blotches of Diaulula sandiegene (Cooper, 1862). Many specimens of Dendronotus 400 have been preserved in Ionol C. P.-40 emulsion and the colors - white, metallic orange, brown, magenta. 1 low, mauve, purple, pink, grey, red - have faded dights over periods of up to 2 years. The orange body color acc white spots of Dirona aurantia Hurst, 1965 and the white pigment on the cerata of D. albolineata CXXXII ELL & ELIOT, 1905 fade somewhat. The dark green but color of Elysia hedgpethi and the black (or dark purph of Aglaja diomedea have faded very little.

In a few cases, the color faded markedly or comparate disappeared. The chocolate brown color of Onehaden bilamellata (Linnaeus, 1767) faded to about a third is original intensity. In Cadlina marginata MacFarlane 1905, the yellow pigment disappears completely with the property of the 24 hours. The yellow on the notal papillar is Acanthodoris hudsoni MacFarland, 1905 fades the Acanthodoris hudsoni MacFarland, 1905 fades the missenda crassicornis and other opisthobranchs.

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Bergin-Wyss, 1961) usually disappear when the sal is killed and they cannot be preserved by an axidant.

CONCLUSION

The use of an antioxidant such as Ionol C. P. -40 with a successful and relatively simple section of preserving most colors in the opisthobranchs.

**Server*, on the basis of this study, it is not possible to server about which colors will be preserved between taxa or even within a certain taxon. Further reperimentation with Ionol C. P. -40 will likely demonstrate a more widespread applicability as well as delinement more precisely which colors will or will not be received.

The optimal concentration proved to be 0.3% Ionol (P.40 by volume in a solution of 5% formalin in sea

Color retention is an obvious advantage to the taxoneast in that it allows him to augment the description of a pecies. This is especially true in soft-bodied animals in opisthobranchs where color, though quite variable straspecifically, often serves as a guide in species identi-

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Laish to extend my thanks to Brian Case, University of Manitoba, for first bringing the method to my attention and to the Shell Chemical Company, Industrial Chemicals Spession (Portland Office) for the sample of Ionol C. P. Lam grateful to Dr. Alan J. Kohn for reading the manuscript and for his suggestions. This work was supported by a Special Scholarship from the National Remarch Council of Canada.

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BOOKS, PERIODICALS, PAMPHLETS

Between Pacific Tides

by Edward F. Ricketts & Jack Calvin, Fourth edition, revised by Joel W. Hedgeeth. September 19, 1968. xiv +614 pp.; 8 color plates; 302 text figures.

Stanford University Press, Stanford, California. \$10.-.

This classic work on the ecology and natural history of plants and animals inhabiting the Pacific shores in the area between the low and the high tide marks was written, originally, "for laymen, for beginners . . ." but it has filled a need far beyond that envisioned by Ed Ricketts. Because of the thoroughly annotated bibliography, the book has become an important first source for many serious investigators and an indispensable text book in many courses in invertebrate zoology.

The book has gone through three previous editions, the first two by the original author, and the third edition revised by Dr. Hedgpeth. Each edition was better than the previous one, each updated to include the latest developments in the fields of endeavor covered by the scope of the work.

This fourth edition, again revised and brought up to date, is a worthy successor to the other three editions. Dr. Hedgpeth has brought his critical talents to bear and his influence can be perceived on practically every page, although he has managed beautifully to preserve the original charm of the book. And while a number of the illustrations are the old familiar ones, many have been superseded by better pictures illustrating more precisely what was desired to be called to the reader's attention. Even the color plates have been much improved, although they may perhaps, in a future edition, be printed with a still better technique which will render the delicate colors of the intertidal area more life-like.